

**Four Year Undergraduate Program (2024-28)**  
**Department of Biotechnology**  
**Course Curriculum**

**Part A: Introduction**

Program: Bachelor in Life Sciences (Diploma/Degree/Honors)		Semester: IV Sem	Session: 2024-2025
1	Course Code	BTSC-04-T	
2	Course Title	Recombinant DNA technology	
3	Course Type	Discipline Specific Course (DSC)	
4	Pre-requisite (if any)	As per program	
6	Course Learning Outcomes (CLO)	After completing this course, the students will be able to - <ul style="list-style-type: none"> <li>• Understand various tools of genetic engineering.</li> <li>• Develop competency in genetic exploitation for human welfare.</li> <li>• Understand the practical application of recombinant DNA technology.</li> <li>• Understand the use of information technology in the field of genome and proteome analysis.</li> </ul>	
6	Credit Value	03 Credits (Credit = 15 Hours - learning & observation)	
7	Total Marks	Max. Marks: 100	Min Passing Marks: 40

**Part B: Content of Course (Theory)**

Total No. of Teaching-learning Periods (01 Hr. per period)- 45 Periods (45 Hours)

Unit	Topic (Course content)	No. of Period
I	<b>Prerequisites of rDNA technology</b> <ol style="list-style-type: none"> <li>1. Recombinant DNA technology: General concept. Steps and application.</li> <li>2. Host controlled Restriction Modification System, Ligases and Polymerases, Klenow fragment, Taq, Pfu polymerase.</li> <li>3. Nuclease (Endo, Exo, and restriction endonuclease).</li> <li>4. Modification Enzyme (Kinase, Phosphates and terminal deoxynucleotidyltransferase), Reverse Transcriptase.</li> </ol>	12 (12 Hrs)
II	<b>Gene transfer</b> <ol style="list-style-type: none"> <li>1. Vectors: Based on Plasmid, Bacteriophages, Cosmid.</li> <li>2. High capacity vectors.</li> <li>3. The basic concept of Gene Transfer Methods: Microinjection, Electroporation, Lipofection, and Microprojectile.</li> <li>4. Selection and Screening of Recombinants: Genetic and Hybridization methods.</li> </ol>	11 (11 Hrs)
III	<b>Genomic validation</b> <ol style="list-style-type: none"> <li>1. PCR: Types of PCR, Steps, Applications, Advantages and Limitations of PCR.</li> <li>2. Molecular Marker-RFLP, RAPD, and Microarray.</li> <li>3. Human Genome Project.</li> <li>4. Gene Library: Genomic and cDNA library, Chromosome walking and jumping.</li> </ol>	11 (11 Hrs)
IV	<b>Application of genetic technology</b> <ol style="list-style-type: none"> <li>1. Gene Therapy: <i>In vivo</i> and <i>Ex vivo</i>, germline and somatic gene therapy.</li> <li>2. Basic idea of stem cell technology: Types of stem cell cultures and their</li> </ol>	11 (11 Hrs)

*[Handwritten signatures and initials in blue ink]*

	Significance. 3. Introduction to Genomics, DNA sequencing methods – manual & automated: Maxam & Gilbert and Sangers method. 4. Introduction to protein structure, Chemical properties of proteins, physical interactions that determine the property of proteins, short-range interactions, electrostatic forces, van der Waal interactions, hydrogen bonds, and Hydrophobic interactions.	
Keywords	Recombinant DNA, Vectors, PCR, cDNA library.	

• Part C - Learning Resource

**Text Books, Reference Books, Other Resources -**

**Text Book-**

- P S Verma and A K Agrawal
- An introduction to genetic engineering- S T Tischoll
- Molecular Biology; Watson.
- Gene VIII; Benjamin Lewin.
- The Cell, A molecular Approach; Geoffrey M. Cooper.
- Molecular Biology of the Cell; Alberts

**Four Year Undergraduate Program (2024-28)**  
**Department of Biotechnology**  
**Course Curriculum**

**Part A: Introduction**

Program: Bachelor in Life Sciences (Diploma/Degree/Honors)		Semester: IV Sem	Session: 2024-2025
1	Course Code	BTSC-04-P	
2	Course Title	Recombinant DNA technology	
3	Course Type	Discipline Specific Course (DSC) - Practical	
4	Pre-requisite (if any)	As per program	
5	Course Learning Outcomes (CLO)	After completing this course, the students will be able to - <ul style="list-style-type: none"> <li>• Isolate nucleic acid from biological cells.</li> <li>• Estimate and manipulate nucleic acid.</li> <li>• Amplify nucleic acid.</li> <li>• Analyse nucleic acid on the basis of database.</li> </ul>	
6	Credit Value	01 Credits	Credit =30 Hours Laboratory or Field learning/Training
7	Total Marks	Max. Marks: 50	Min Passing Marks: 20

**Part B: Content of Course**

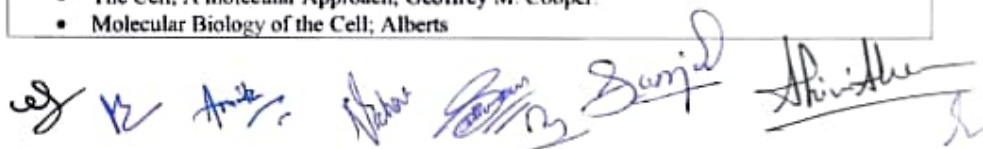
Total No. of learning-Training/performance Periods: 30 Periods (30 Hours)		
Module	Topic (Course content)	No. of Period
Lab./Field Training/ Experiment Contents of Course	<ol style="list-style-type: none"> <li>1. Isolation of chromosomal DNA from plant cells</li> <li>2. Isolation of chromosomal DNA from <i>E.coli</i></li> <li>3. Qualitative and quantitative analysis of DNA using spectrophotometer</li> <li>4. Plasmid DNA isolation</li> <li>5. Restriction digestion of DNA.</li> <li>6. Ligation of DNA.</li> <li>7. Transformation of competent cells.</li> <li>8. Demonstration of PCR.</li> <li>9. Use of SNP databases at NCBI and other sites.</li> <li>10. Use of OMIM database</li> <li>11. Detection of Open Reading Frames using ORF Finder</li> </ol>	30
Keywords	Recombinant DNA, Vectors, PCR, cDNA library.	

**Part C - Learning Resource**

**Text Books, Reference Books, Other Resources -**

**Text Book-**

- P S Verma and A K Agrawal
- An introduction to genetic engineering- S T Tischoll
- Molecular Biology; Watson.
- Gene VIII; Benjamin Lewin.
- The Cell, A molecular Approach; Geoffrey M. Cooper.
- Molecular Biology of the Cell; Alberts



- Modern Biotechnology, 2nd Edition, S.B. Primrose, Blackwell Publishing, 1987.
- Molecular Biotechnology: Principles and Applications of Recombinant DNA, 4th Edition, B.R. Glick, J.J. Pasternak and C.L. Patten, 2010.
- Molecular Cloning: A Laboratory Manual (3rd Edition) Sambrook and Russell Vol. I to III, 1989.